

## SYNTHESIS OF INORGANIC PYROPHOSPHATE BY ANIMAL TISSUE MITOCHONDRIA

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### 1. Introduction

It is known that inorganic pyrophosphate ( $PP_i$ ) is formed as a result of pyrophosphorolysis reactions (synthesis of nucleic acids, coenzymes, proteins, activation of fatty acids etc.). It was believed that, to shift the equilibrium of these reactions towards synthesis,  $PP_i$  should be hydrolyzed by inorganic pyrophosphatase, the energy of anhydride bonds being thereby spent [1].

But some facts have been accumulating, testifying that  $PP_i$  in some biochemical conversions may perform a function similar to that of ATP [2–8]. In 1942 Cori [9] reported to have observed accumulation of  $PP_i$  in oxidation of glutamate by rat liver homogenates and Green et al. [10], in 1949, by rabbit liver homogenates. Synthesis of  $PP_i$  as a result of oxidative phosphorylation was demonstrated to take place in *Acetobacter suboxidans* and yeast [11] and as a result of photophosphorylation, in *Rhodospirillum rubrum* [12].

The aim of this work was to elucidate the possibility of formation of  $PP_i$  and of its utilization for the ATP synthesis in rat liver mitochondria.

### 2. Materials and methods

Mitochondria were isolated by means of a modified method of Schneider [13]. Their Lardi-Welman respiration coefficient was 3.0 to 3.5. ATP synthesis was determined in an acid extract of mitochondria with hexokinase and glucose 6-phosphate dehydrogenase [14]. To avoid possible formation of ATP from ADP with participation of myokinase,

the incubation medium was supplemented with AMP. Therefore, to compare the rates of  $PP_i$  synthesis and phosphorylation with the participation of the adenylic system, not only ATP increment but also ADP formation were to be taken into consideration. To determine ADP, it was converted into incubating it with PEP and pyruvate-kinase [15].  $PP_i$  was determined colorimetrically in the presence of  $\beta$ -mercaptoethanol and metabisulphite after three-fold extraction of orthophosphate in the form of a phosphomolybdate complex [16]. Inorganic orthophosphate was determined by the method of Behrenblum and Chain as modified by Weil-Malherbe and Green [17]. To identify  $PP_i$  it was precipitated in the form of barium salts from the acid extract of mitochondria and subjected to t.l.c. on DEAE cellulose and paper.

### 3. Results and discussion

As is shown in fig.1, rat liver mitochondria in the oxidative phosphorylation conditions synthesize, in addition to ATP,  $PP_i$ . A similar synthesis was detected in beef heart mitochondria. In our experiments, the content of ADP exceeded that of ATP on an average 3–4 fold, and phosphorylation involving the adenylic system was 5–10 times as intensive as the  $PP_i$  synthesis.

One could suggest that  $PP_i$  synthesis is associated with the functioning of mitochondrial electron transport chain. To verify this, we have studied the effect on the  $PP_i$  biosynthesis of 2,4-dinitrophenol, an uncoupler, and electron transport inhibitors at the level of the second and third coupling sites. As is seen in fig.2, dinitrophenol ( $4 \times 10^{-4}$  M), antimycin ( $2 \mu\text{g}/\text{mg}$  protein) and NaCN ( $10^{-3}$  M) inhibited  $PP_i$



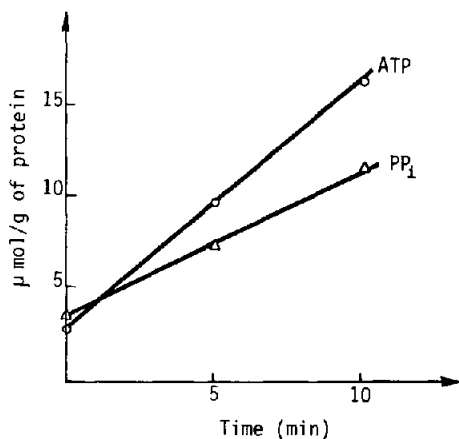


Fig. 1. Synthesis of ATP and  $PP_i$  in rat liver mitochondria. Mitochondria were incubated at room temperature in the medium of the following composition: sucrose—0.25 M, Tris—HCl buffer pH 7.4—0.005 M, succinate—0.037 M, AMP—0.003 M, EDTA—0.0008 M,  $KH_2PO_4$ —0.0075 M,  $MgSO_4$ —0.03 M. The volume of the incubation medium—2 ml, protein—10–15 mg/ml. The reaction was interrupted by perchloric acid.

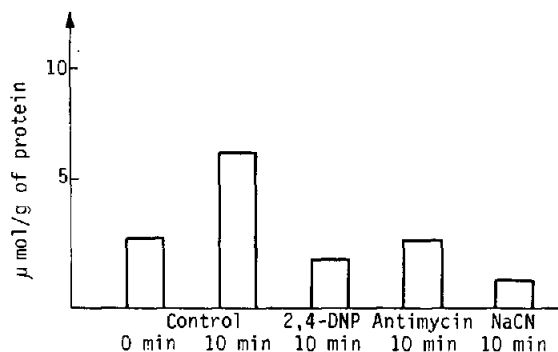


Fig. 2. Effect of an uncoupler and respiratory chain inhibitors on  $PP_i$  synthesis. Incubation conditions are the same as in fig. 1.

formation. In the mitochondria with uncoupled oxidation and phosphorylation (incubation in a sucrose-less medium), these compounds produced no effect on the pyrophosphatase and ATPase activities of mitochondria. The results obtained indicate that the synthesis of pyrophosphate is associated with the functioning of electron transport chain.

Two possible pathways of  $PP_i$  formation may be

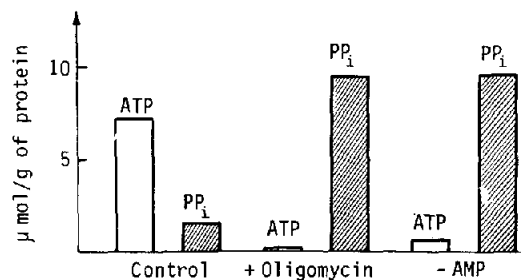


Fig. 3. Synthesis of ATP and  $PP_i$  in the presence of oligomycin. Incubation conditions are the same as in fig. 1. Time of incubation—10 min.

inferred from the above data. The first is hydrolysis of ATP to AMP and  $PP_i$ , and the second—an ATP-independent pathway. To choose between the two possibilities, the effect of oligomycin, and ATPase inhibitor, on  $PP_i$  synthesis was studied.

As is shown in fig. 3, the action of oligomycin (1  $\mu$ g/mg mitochondrial protein) induces stimulation of  $PP_i$  synthesis with the synthesis of ATP being inhibited. Similar results were obtained without AMP being added to the incubation medium. This testifies in favour of an independent formation of  $PP_i$ . The data obtained are in agreement with those of H. Baltscheffsky [12] on the effect of oligomycin on the  $PP_{in}$  synthesis in chromatophores. The data presented in fig. 4 confirm this pathway. Preincubation of mitochondria in the presence of oligomycin, hexokinase and glucose in the absence of phosphate brought about disappearance of ATP and ADP and stabilization of the  $PP_i$  level. Further addition of phosphate stimulated ATP-independent  $PP_i$  synthesis.

In the laboratories of Baltscheffsky and Keister [1,3] it was demonstrated that inorganic pyrophosphatase is involved both in the biosynthesis and utilization of  $PP_i$ . We have detected inorganic pyrophosphatase to be present in a highly purified fraction of rat liver mitochondria. The activity was calculated per unit of cytochrome  $a$  which is known to be localized in mitochondria, and it was demonstrated that the pyrophosphatase activity of the cell is mostly localized in mitochondria. Electrophoresis in polyacrylamide gel revealed two isoenzymes to be present, of which at least one is firmly bound to the membrane. Similar data were previously reported by H. Baltscheffsky et al. [12] for pyrophosphatase



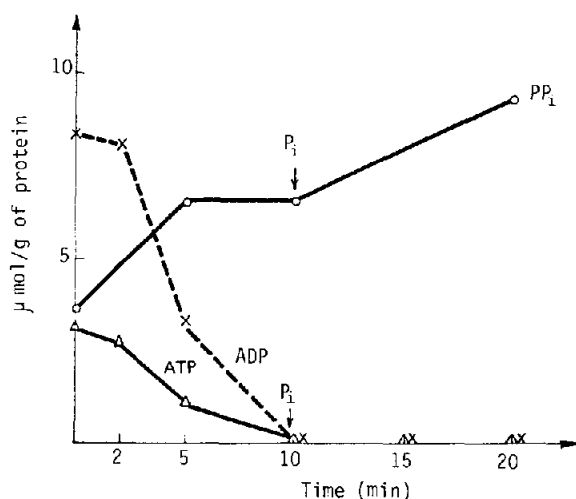


Fig. 4.  $PP_i$  synthesis by ADP and ATP-exhausted mitochondria. The mitochondria were preincubated at room temperature with hexokinase (0.1 mg/ml medium), oligomycin (1  $\mu$ g/mg protein) and glucose ( $4 \times 10^{-2}$  M) in the medium of fig. 1 but in the absence of phosphate. In 10 min  $KH_2PO_4$  - 0.0025 M, was added.

from *R. rubrum* chromatophores and by M. Baltscheffsky [18] and Iria et al. [19] for pyrophosphatase from rat liver mitochondria.

A comparative analysis of the effect of various inhibitors on phosphatase and ATPase of mitochondria has shown that respiratory chain inhibitors (antimycin, sodium cyanide) produces no effect whatsoever of ATPase and pyrophosphatase. Oligomycin inhibited only ATPase and sodium fluoride at a concentration of

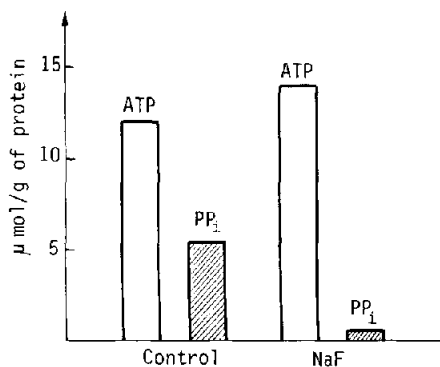


Fig. 5. Effect of sodium fluoride on the mitochondrial ATP and  $PP_i$  synthesis. Incubation medium is the same as in fig. 1. Concentration of NaF -  $10^{-2}$  M. Time of incubation - 10 min.

$10^{-2}$  M preferentially inhibited pyrophosphatase. It should be noted that NaF inhibits both ATPase and pyrophosphatase. But with  $10^{-2}$  M NaF and 10–15 mg/ml mitochondrial protein pyrophosphatase is inhibited preferentially and ATPase - to a small extent. The pattern of inhibition of these two enzymes seems to be different. Our preliminary data may point to the fact that in the case of pyrophosphatase NaF not only inhibits the enzyme but also affects its interaction with the membrane. As is shown in fig. 5, incubation of mitochondria in the conditions of oxidative phosphorylation with NaF at a concentration of  $1 \times 10^{-2}$  M selectively inhibits the synthesis of  $PP_i$ . The ATP synthesis is not thereby inhibited, it is even stimulated. These data should be interpreted as favouring the participation of pyrophosphatase of mitochondria in  $PP_i$  biosynthesis. It is appropriate to mention that Racker's coupling factor  $F_3$  proved to display a pyrophosphatase activity [20].

Our data on the effect of oligomycin and NaF on the ATP and  $PP_i$  synthesis indicated that a common intermediate is involved in the biosynthesis of these compounds and the energy of electron transport may be consumed either for the synthesis of ATP or for that of  $PP_i$ .

To verify the possibility of ATP biosynthesis at the expense of  $PP_i$ , a series of experiments was performed in which the change in the ATP content in mitochondria was followed up during their incubation with and without  $PP_i$ . The relevant results are shown in fig. 6. As is clear from the figure, in the presence of  $PP_i$  the intensity of ATP biosynthesis markedly increases. Special experiments were designed to demonstrate that an increased concentration of orthophosphate in the incubation medium did not affect the increment of ATP, and it was only the presence of  $PP_i$  that entailed an increase in the ATP concentration in the experimental conditions employed.

The respiratory chain inhibitor (NaCN) almost completely inhibited the ATP synthesis at the expense of oxidative phosphorylation. Addition of  $PP_i$  to the incubation medium in the same conditions led to an ATP synthesis. It is of interest that the difference in the amount of ATP synthesized in the presence and absence of  $PP_i$  and the quantity of ATP synthesized in the presence of  $PP_i$  and sodium



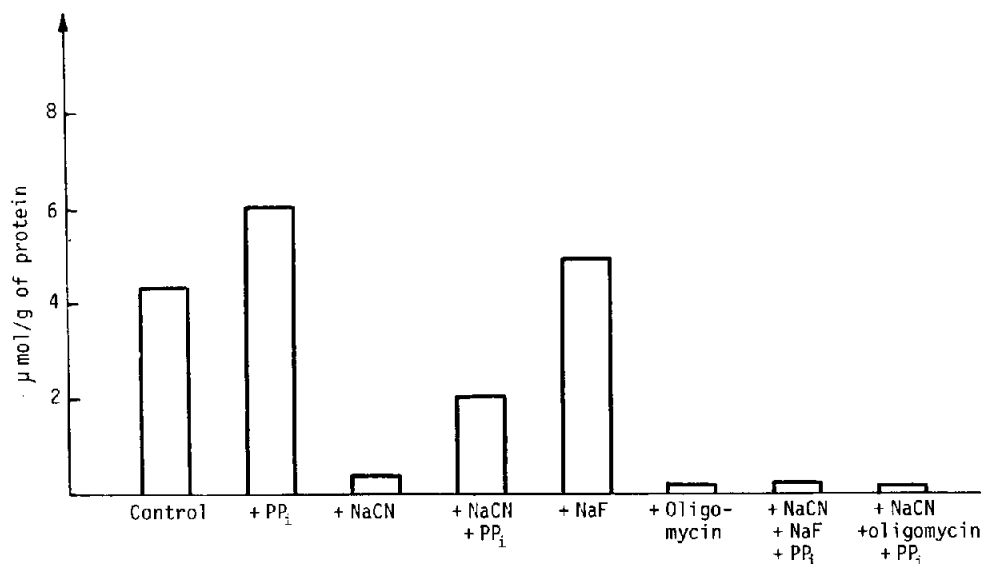
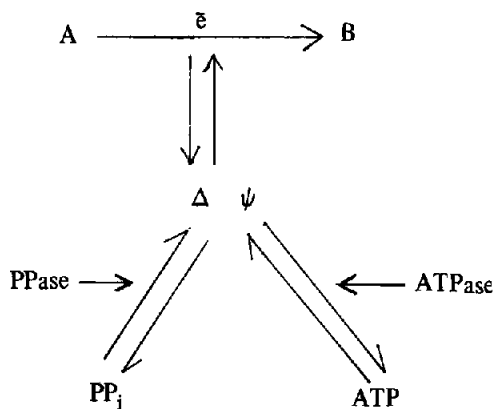


Fig.6. PP<sub>i</sub>-dependent ATP synthesis. Incubation medium is the same as in fig.1. Concentration of added PP<sub>i</sub>— $1 \times 10^{-3}$  M, NaCN— $1 \times 10^{-3}$  M, NaF— $1 \times 10^{-2}$  M, oligomycin— $2 \mu\text{g}/\text{mg}$  protein. Time of incubation—10 min.

cyanide, i.e. at the expense of PP<sub>i</sub> only, are rather close.

With the use of NaF and oligomycin against the background of cyanide, no PP<sub>i</sub>-dependent ATP increment was observed. This means that in the PP<sub>i</sub>-ATP reaction inorganic pyrophosphatase and ATPase are involved. There are grounds for believing that utilization of the PP<sub>i</sub> energy for ATP synthesis is a process similar to that in *Rhodospirillum rubrum* chromatophores which proceeds according to the following scheme:



in which membrane electric potential should be the common source of energy [8,21].

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